

Original Research Article

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## Genetic Diversity and Principal Component Analysis for Grain Quality and Productivity Traits in Sorghum [*Sorghum bicolor* (L.) Moench]

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### ABSTRACT

An investigation was carried out to assess nature and magnitude of genetic diversity for grain quality traits and productivity traits in mini core collection of sorghum. Mini core accessions were grouped into 15 clusters, where in cluster III had largest with 21 accessions whereas cluster XIII had minimum with 5 accessions. Plant height contributed maximum divergence with 46.2%. The inter cluster distance  $D^2$  value ranged widely with minimum values of ( $D^2=197.61$ ) and maximum value ( $D^2=5541.42$ ) indicating high diversity among mini core and it was desirable to select mini core from clusters showing high inter cluster distance. Diversity among cluster varied from ( $D^2=255.25$ ) to ( $D^2=4906.5$ ) inter cluster distance. Principal component analysis revealed that, three out of nine principal components with eigenvalues  $> 1$  were extracted. These three components contributed 58.29% of the total variation among the mini core. Principal components first three contributed, 22.73%, 17.99%, and 15.50%, respectively toward the variation observed among accessions. Variation relative to the first component was associated with seed yield per plant, 100 seed weight, seed volume, bulk density, seed size. The second principal component was associated with plant height, ear head length, ear head width, seed yield per plant, 100 seed weight, seed volume and seed size. The third principle component was associated with ear head width, 100 seed weight, seed yield per plant and seed size.

#### Keywords

Sorghum, Diversity, Principal component, Mini core, Cluster

#### Article Info

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### Introduction

Sorghum [*Sorghum bicolor* (L.) Moench] is one of the important cereal crop in the world, which is grown in Africa, Asia, USA, Australia and Latin America. It is the fourth most important cereal crop following rice, wheat, maize and staple food in the same central parts of the world. Worldwide, it is

cultivated on 41.07 million ha area with production of 58.42 million tones in the year approx., 2019-20 (Anonymous 2019a). In India, sorghum having 5.00 million ha area with 4.5 million tones production and 900 kg/ha productivity in the year 2019-20 (Anonymous 2019b). Sorghum shows extreme genetic diversity (Sanchez *et al.*, 2002) and is predominantly self-pollinating, with varying

levels of outcrossing. Sorghum grown in *rabi* season is characterized by its excellent grain quality, exclusively utilized for human consumption and hence fetches higher market price as compared to *kharif*.

Understanding of genetic diversity of a species is fundamental in any crop improvement programme. For such species, in general the parents with more genetic divergence are expected to yield heterotic hybrids in addition to generating a broad spectrum of variability in segregating generations. The  $D^2$  statistic is a useful multivariate statistical tool for effective discrimination among various genotypes on the basis of genetic divergence (Murty and Arunachalam, 1966; Sonawane and Patil, 1991). Diversity analysis provides information on deciding choice of parents from distantly related clusters to secure yield improvement in sorghum. A better understanding of genetic diversity in sorghum will facilitate crop improvement (Jayarama Chandran *et al.*, 2011). Diversity in germplasm is important for any breeding program, since it directly affects the potential for genetic gain through selection (Kotal *et al.*, 2010). Genetic diversity among the genotypes serves as a way to adapt to changing environments and their utilization in crop improvement programme. However, reports on genetic diversity among the *rabi* sorghum is very limited. Therefore there is a need to evaluate the available accessions for genetic diversity.

Principal component analysis is a multivariate technique for examining the relationships among several quantitative variables (Johnson 2012). It is the most common technique used in variability studies and numerical classification; it is useful in grouping varieties based on their similarities (Bello 2004). Principal component analysis is an important breeding tool commonly used by breeders to identify traits that could be used to discriminate crop genotypes (Das 2000; Yan

and Kang 2003). Establishing suitable selection criteria for identifying genotypes with desirable traits is useful in developing improved varieties. Analysis of variability among traits and knowledge of associations among traits contributing to yield would be of great importance in planning a successful breeding program (Mary and Gopalan 2006). To date, in Niger, no study has been carried out with the objective to assess diversity in sorghum based on traits mentioned above by using multivariate analysis. Therefore, the objective of this study is to determine genetic diversity of sorghum inbred lines, which would be helpful in enhancing the efficiency of sorghum breeding program.

Suitable selection criteria for the identification of genotypes with desirable traits are essential for successful varietal improvement programs. Analysis of variability among traits and the identification of associations among various traits contributing to yield would facilitate successful development of high yielding varieties (Mary and Gopalan 2006). However, selecting only for grain yield may not be efficient for developing varieties for adoption by farmers; selection, which integrates yield and farmer-preferred traits, should provide more appropriate varieties (Alvi *et al.*, 2003). The identification of yield-related traits could result in more effective selection for yield and farmer-preferred traits. The high level of genetic diversity and characterization of accessions integrated into world collections is essential in order to classify, manage exotic germplasm, collect and ultimately utilize the different genetic improvement of the crop.

## **Materials and Methods**

The present investigation was carried out during *rabi* season 2011-12 at AICSIP, UAS, Dharwad. The plant material for this experiment comprised of 208 accessions of mini core collection obtained from DSR

Hyderabad. The experiment was laid out in medium deep black soil under rain fed condition. The randomized block design was followed separately with two replications and each entry was sown in four rows of 4 m length with inter row spacing of 45 cm and intra row spacing of 15 cm. Observations on all quantitative characters like plant height (cm), panicle length (cm), panicle width (cm), seed yield per plot (g), 100 seed weight (g), seed volume (ml), bulk density (g/ml), true density (g/ml) and seed size (mm).

Seed size was measured by using Vernier Callipers where length, breadth and thickness of seeds were recorded. Seed density classified into two types *viz.*, true density and bulk density. Seed bulk density was measured by hundred gram of seeds were weighed and volume was recorded in a measuring jars. Whereas, seed true density was observed by known weight of seeds placed in a measuring jar containing known quantity of toluene. Increase in volume was recorded after pouring seeds in measuring jar. Seed volume was noted with countable numbers of seeds were placed in a measuring jar. Grain quality characters like seed luster, seed color, seed shape and seed hardness was recorded by measuring the grinding time required to obtain a fixed volume of flour from the grains. Mean of five plants for each entry was worked out and used for statistical analysis. Genetic diversity was studied using Mahalanobis  $D^2$  statistic and clustering was done following Tocher's method described by Rao (1952) for determining group constellation. Average inter and intra cluster distances were estimated as per the procedure outlined by Singh and Choudhary (1977).

The analysis of variance for the individual character and analysis of covariance for character pairs were carried out as described by Cochran and Cox (1957). Divergence was estimated by the multivariate analysis using

Mahalanobis's (1936) and  $D^2$  statistic as described by Rao (1952). On the basis of  $D^2$  values obtained, the variables were grouped into different clusters by employing Tocher's method (Rao, 1952). The percent contribution of each character to the total divergence was calculated by ranking each character on the basis of transformed uncorrelated values. Finally, the percent contribution for each character was calculated by taking total number of ranks of all the characters to hundred. The data were analyzed statistically using the software WINDOSTAT, developed by INDOSTAT services Ltd. Hyderabad, India.

## **Results and Discussion**

The analysis of variance showed highly significant differences among the accessions for all the characters studied indicating the presence of considerable variability in the experimental material. Nature and magnitude of genetic diversity exists in the crop species will be utilized for formulating breeding programme. Mahalanobis'  $D^2$  statistics is used to quantify the degree of divergence. It is based on second degree statistics and pattern obtained by  $D^2$  does not change with number of characters. Based on  $D^2$  statistics and tocher method 208 accessions were grouped into 15 clusters with variable number of entries revealing the presence of considerable amount of genetic diversity in the material. Among them, cluster III had largest with 21 accessions whereas cluster XIII had minimum with 5 accessions reflecting narrow genetic diversity among them. Cluster VIII with 19 accessions, Whereas, three cluster namely IV, V and XIV had 17 accessions followed by Cluster VI and X with 16 accessions, cluster I and XV with 15 accessions, cluster II and IX had 12 accessions each. Whereas, cluster VII had 11 accessions, cluster XI had 8 accessions and cluster XII had 7 accessions, respectively (Table-1). The narrow genetic diversity may

be attributed to similarity in the base material from which they have been evolved.

Among nine quantitative traits studied, the highest contribution towards the divergence was by plant height (46.2%). Similar results were reported by Kukadia *et al.*, (1981), Sisodia *et al.*, (1983), Dabholkar *et al.*, (1983) and Mehendiratta and Sindhy (1972). Interestingly grain quality traits like seed size contributed 23.49% followed by bulk density (12.03%), seed volume (6.65%), true density (5.44%) including seed yield per plant (5.77%), which indicates that grain quality traits also contributing for diversity. However, the characters like ear head width (0.39%), ear head length (0.02%) and 100 seed weight (0.01%) indicated narrow range of diversity among the mini core under study (Table-2).

The average intra (diagonal) and inter cluster (off diagonal)  $D^2$  values are presented in the table-5. The inter cluster distance  $D^2$  value ranged widely with minimum values of ( $D^2=197.61$ ) and maximum value ( $D^2=5541.42$ ) indicating high diversity among mini core and it was desirable to select mini core from clusters showing high inter cluster distance. Diversity among cluster varied from ( $D^2=255.25$ ) to ( $D^2=4906.5$ ) inter cluster distance (Table-3). Higher intra cluster distance indicates that genotypes in the respective clusters and the higher inter cluster distances have wider genetic distances between the genotypes which could be used in hybridization programme.

In the present investigation, the inter cluster distance was higher than intra cluster distance which indicated substantial diversity among the mini core accessions and there may be a greater opportunity for obtaining the rare but superior segregants from crosses between more divergent accessions. Similar results were also obtained by earlier investigators (Swami *et al.*, 2015; Jain and Patel, 2013; and Mohanraj *et al.*, 2006).

The maximum inter cluster distance observed was between cluster XI and XIII (5541.42) followed by cluster XI and XII (4942.89), cluster IX and XIII (4225.12) and cluster VII and XI (4225.12). Intra cluster distance  $D^2$  ranged from 0 to 596.24 which was followed by cluster V ( $D^2=570.19$ ). The most of intra cluster distance was zero. The intra cluster distance  $D^2$  value ranged widely with minimum value of 0 were observed between most of the clusters followed by (230.13) cluster I and I and cluster I and II was (247.31).

Cluster mean analysis was calculated using Tocher's method for nine yield and its attributing traits and presented in Table 4. Higher cluster mean for plant height was observed in cluster XI (292.68) followed by cluster VIII (263.67) and cluster IX (263.34). Whereas, lower cluster mean was recorded in cluster XIII (98.17). For ear head length cluster mean was recorded in cluster IX (35) followed by cluster XI (32.92) and cluster X (31.27). However, lower cluster mean in cluster VII (19.33). For earhead width the highest cluster mean was recorded in cluster X (13.33) and lowest were recorded in cluster I, cluster VII and cluster XII (7.5). Highest and lowest cluster mean for seed yield per plant was recorded in cluster XIV (42.5) and cluster VIII (5.95), respectively.

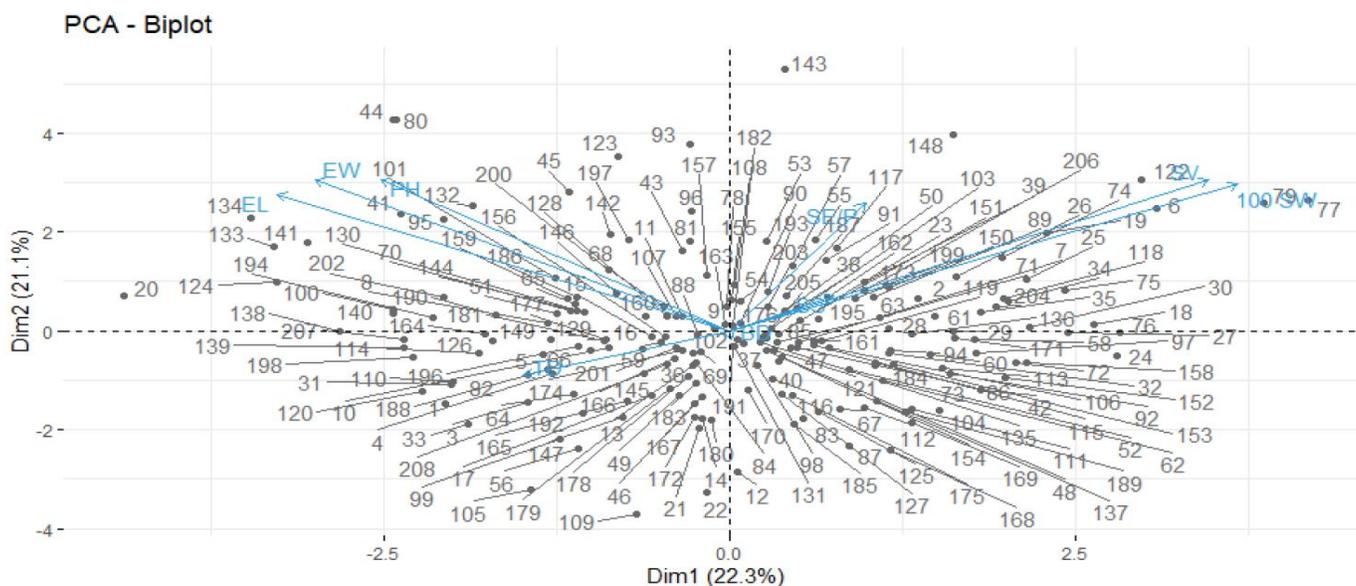
Based on overall score across nine traits, the cluster were ranked. Accordingly, cluster XIII with overall scores of 38 across XV clusters secured first rank followed by cluster XII, cluster VII, cluster I and cluster IV are the top clusters, indicating the presence of most promising accessions in them and can be extensively used for further breeding programme to generate new material.

The purpose of principal component analysis is to reduce the volume of data. Watson and Eyzaguirre (2002) also reported that PCA of morphological characterization results could

identify a few key or minimum descriptors that effectively account for the majority of the diversity observed, saving time and effort for future characterization efforts. Principal components approach is very helpful in

deciding which agronomic traits of crop contributing most to yield, subsequently, these agronomic traits should be emphasized in the breeding program (Jain *et al.*, 2016).

**Figure.1** The mini core accession by trait biplots of *rabi* sorghum



**Table.1** Per cent contribution of characters towards divergence 208 mini core collections of *rabi* sorghum

SI	Source	Contribution %
1	Plant height	46.2°/
2	Ear head length	0.02°/
3	Ear head width	0.39°/
4	Seed yield per plant	5.77°/
5	100 seed weight	0.01°/
6	Seed volume	6.65%
7	Bulk density	12.03%
8	True density	5.44%
9	Seed size	23.49°/

**Table.2** Distribution of 208 mini core collections of *rabi sorghum* into different cluster

Cluster No	No. of mini core	Within SS	Cluster members
1	15	0.6330	IS-602, IS-1233, IS-2389, IS-2413, IS-2426, IS-3971, IS-4060, IS-4951, IS-8012, IS-9177, IS-24453, IS-26749, IS-29714, IS-30572, IS-33353
2	12	0.7812	IS-473, IS-1004, IS-4515, IS-6351, IS-10302, IS-10757, IS-12302, IS-13893, IS-14779, IS-25089, IS-27034, IS-28449
3	21	1.2792	IS-1041, IS-2864, IS-4360, IS-4698, IS-6354, IS-8916, IS-9108, IS-12735, IS-12883, IS-14010, IS-15466, IS-15744, IS-24953, IS-29241, IS-15466, IS-15744, IS-24953, IS-29241, IS-29269, IS-29565, IS-29568, IS-29606, IS-29654, IS-30383, IS-30443
4	17	1.5104	IS-2382, IS-7131, IS-305, IS-11919, IS-13782, IS-16382, IS-19153, IS-19445, IS-28849, IS-29239, IS-29468, IS-29914, IS-30079, IS-30417
5	17	2.1486	IS-995, IS-10867, IS-13294, IS-13549, IS-25910, IS-25989, IS-26222, IS-27887, IS-29233, IS-29392, IS-29304, IS-29733, IS-30092, IS-30400, IS-30838, IS-31043, IS-31557, IS-33023
6	16	0.4695	IS-1219, IS-4631, IS-5094, IS-5301, IS-6421, IS-13971, IS-14290, IS-15478, IS-18038, IS-25732, IS-26737, IS-29187, IS-29627, IS-30451, IS-30507, IS-31651
7	11	0.6991	IS-4092, IS-12447, IS-14090, IS-19676, IS-24348, IS-24462, IS-27912, IS-28141, IS-29358, IS-29392, IS-29582
8	19	3.4635	IS-20298, IS-20679, IS-20697, IS-20727, IS-21512, IS-21645, IS-21863, IS-22239, IS-22609, IS-22720, IS-22986, IS-23514, IS-23521, IS-23579, IS-23583, IS-23590, IS-23684, IS-23891
9	12	5.4003	IS-20625, IS-20632, IS-20740, IS-20743, IS-21083, IS-22294, IS-22626, IS-23216, IS-23992, IS-24139
10	16	0.8129	IS-603, IS-608, IS-995, IS-1212, IS-5295, IS-5919, IS-12945, IS-19389, IS-24939, IS-25548, IS-26694, IS-29314, IS-30460, IS-30536, IS-31186
11	8	0.7865	IS-7987, IS-15931, IS-15945, IS-19975, IS-26025, IS-26484, IS-28451, IS-28614
12	7	1.4604	IS-7250, IS-7310, IS-7679, IS-25242, IS-25301, IS-26046, IS-28747
13	5	0.8236	IS-2397, IS-2872, IS-3158, IS-19262, IS-29950
14	17	0.6332	IS-2379, IS-4581, IS-4613, IS-6421, IS-9113, IS-12937, IS-16151, IS-17980, IS-24463, IS-24492, IS-26701, IS-29326, IS-29335, IS-29689, IS-29772, IS-30450
15	15	0.9438	IS-2902, IS-8774, IS-12706, IS-13919, IS-14861, IS-15170, IS-19450, IS-19859, IS-25836, IS-28313, IS-29441, IS-29519, IS-30538, IS-31714

**Table.3** Average D<sup>2</sup> values of intra and inter cluster distances among 208 mini core collections of *rabi sorghum*

Cluster	Cluster I	Cluster II	Cluster III	Cluster IV	Cluster V	Cluster VI	Cluster VII	Cluster VIII	Cluster IX	Cluster X	Cluster XI	Cluster XII	Cluster XIII	Cluster XIV	Cluster XV
Cluster I	230.13	583.66	516.19	1053.52	459.46	455.31	949.28	1225.29	1126.62	1561.84	1693.2	1221.3	1468.32	1381.21	829.85
Cluster II		247.31	1449.25	1362.04	785.64	1026.81	449.02	2191.09	2428.62	2811.28	3330.21	373.93	575.67	1799.39	1299.95
Cluster III			323.5	1417.27	760.53	599.27	2008.31	1034.7	545.11	1121.36	886.14	2504.93	2815.12	1690.87	947.62
Cluster IV				373.65	1442.88	1231.76	1731.23	602.41	1944.02	1067.37	2893.72	1768.22	1910.46	649.15	1922.73
Cluster V					5770.19	839.51	1162.22	1690.12	1350.38	1930.15	1832.08	1522.88	1881.72	1509.75	1040.33
Cluster VI						260.9	1080.54	1072.24	1355.03	1615.05	1811.89	1812.28	1687.81	1526.97	1598.48
Cluster VII							0	2602.62	3561.47	3575.37	4225.12	601.67	435.73	1783.18	2462.68
Cluster VIII								0	1142.4	509.4	1931.98	3113.22	3166.75	900.84	2131.91
Cluster IX									0	1053.95	629.13	3725.28	4290.56	2460.67	1033.13
Cluster X										480.5	1521.27	3883.24	4247.78	1244.99	1945.98
Cluster XI											596.24	4942.89	5541.42	2982.2	1650.1
Cluster XII												0	197.61	2567.58	1820.74
Cluster XIII													0	2638.1	2710.63
Cluster XIV														0	2871.46
Cluster XV															0

**Table.4** Clusters means of 9 quantitative characters among 208 mini core collections of *rabi sorghum*.

Cluster No	Plant Height	Ear head length	Ear head width	Seed yield per plant	100 seed weight	Seed volume	Bulk Density	True Density	Seed size
<b>I</b>	190.55	20.76	7.5	27.17	2.49	0.04	0.71	1.26	3.33
<b>II</b>	136.74	21.7	6.88	25.44	2.5	0.04	0.73	1.29	3.49
<b>III</b>	234.52	26.15	9.88	28.29	2.28	0.04	0.7	1.29	3.27
<b>IV</b>	176.06	21.66	7.57	27.13	2.58	0.04	0.71	1.31	0.03
<b>V</b>	191.82	21.59	7.99	30.75	3.06	0.05	0.74	1.3	3.7
<b>VI</b>	202.19	25.1	10.05	27.25	2.04	0.03	0.56	1.2	3.27
<b>VII</b>	122.17	19.33	7.5	15.15	3.56	0.06	0.59	1.1	3.74
<b>VIII</b>	263.67	28.33	8.5	5.95	1.47	0.04	0.63	1.42	0.02
<b>IX</b>	263.34	35	8.34	27.7	1.97	0.03	0.77	1.64	2.89
<b>X</b>	249.87	31.27	13.33	33.36	2.62	0.04	0.75	1.29	0.03
<b>XI</b>	292.68	32.92	11.19	29.93	2.66	0.04	0.76	1.28	3.58
<b>XII</b>	97.17	20	4.84	25.75	2.39	0.03	0.77	1.24	3.1
<b>XIII</b>	89.17	25.84	7.5	28.25	1.75	0.03	0.63	1.24	3.03
<b>XIV</b>	188.33	21.5	10.5	42.5	4.23	0.09	0.67	1.24	0.04
<b>XV</b>	202.83	27.5	8.84	13.05	1.78	0.01	0.95	1.27	3.55

**Table.5** Principal component analysis of measured traits in 208 mini core accessions of *rabi* sorghum

	Plant height	Earhead length	Earhead width	Seed yield per plant	100 seed weight	seed volume	Bulk density	True density	Seed size
<b>eigenvalue</b>	2.01	1.9	1.34	1.05	0.9	0.63	0.55	0.43	0.2
<b>variance.percent</b>	22.31	21.12	14.86	11.66	10	7.03	6.1	4.75	2.18
<b>cumulative.variance.percent</b>	22.31	43.43	58.29	69.94	79.95	86.98	93.07	97.82	100

**Table.6** Factor loadings of the study traits of the first three principal components (PCs)

Traits	Mini core		
	PC1	PC2	PC3
<b>PH</b>	-0.3390	0.4228	-0.0093
<b>EL</b>	-0.4392	0.3813	-0.1143
<b>EW</b>	-0.4030	0.4218	0.1124
<b>SE/P</b>	0.4949	0.4089	-0.2060
<b>100 SW</b>	0.1322	0.3566	0.0709
<b>SV</b>	0.4660	0.4217	-0.0953
<b>BD</b>	0.0022	-0.0248	-0.7121
<b>TD</b>	-0.1997	-0.1234	-0.6403
<b>SS</b>	0.1015	0.0997	0.0252

A screen plot is a simple line segment plot that shows the fraction of total variance in the data. It is a plot, in descending order of magnitude, of the eigen values of a correlation matrix. According to Chatfield and Collins (1980), components with an eigenvalue of  $<1$  should be eliminated so that fewer components are dealt with. Sharma (1998) reported that PCA reflects the importance of the largest contributor to the total variation at each axis of differentiation. It was further reported by Fenty (2004) that PCA reduces a large set of variables to come up with smaller sets of components those summaries the correlations. The Screen plot of the PCA (Fig. 1) shows that the first three eigenvalues correspond to the whole percentage of the variance in the dataset.

Three out of nine principal components with eigenvalues  $> 1$  were extracted. These three components contributed 58.29% of the total variation among the germplasm. Principal components 1, 2, and 3 contributed, respectively, 22.73%, 17.99%, and 15.50% toward the variation observed among genotypes (Table-5). The aim of principal component analysis is to resolve the total variation of a set of traits into linear, independent composite traits, which successively maximize variability in the data (Johnson 2012). Sample traits are generally inter-correlated to varying degrees and hence not all principal components are needed to summarize the data adequately. In this study, the first three principal components represented a sizeable amount of diversity among the genotypes investigated. This implied that several traits were involved in explaining the variation among the genotypes. Ayana and Bekele (1999) reported significance of first five PCs in the total variability of different agro-morphological traits in sorghum. The first four principal components, with eigenvalues greater than one, were also documented in 25 forage and

45 grain sorghum genotypes for dual purpose (Chikuta *et al.*, 2015). Abraha *et al.*, (2015) reported four principal components with eigenvalues greater than one, which explained  $> 75%$  of the total variation for grain yield, biomass, stay-green, leaf area, peduncle exertion, days to flowering, and maturity. Around 44%, 17%, and 15% variation attributed to first, second, and third principal components, respectively, was reported by Chikuta *et al.*, (2015). Several studies on principal component analysis of different agro-morphological traits in sorghum have been documented. Abraha *et al.*, (2015) concluded that grain yield, biomass, stay-green, leaf area, peduncle exertion, days to flowering, and maturity were the most important traits for genetic variability in landrace sorghums. On the other hand, head width, head weight, grain yield per plant, and fresh and dry shoot weight were found to be the most important traits for drought tolerance in grain sorghum (Ali *et al.*, 2011).

The phenotypic diversity observed in this study was attributable to several traits (Table-6). Variation relative to the first component was associated with seed yield per plant, 100 seed weight, seed volume, bulk density, seed size. The second principal component was associated with plant height, ear head length, ear head width, seed yield per plant, 100 seed weight, seed volume and seed size. The third principle component was associated with ear head width, 100 seed weight, seed yield per plant and seed size. Distribution of biometrical traits in first two components is shown in loading plot (Fig. 1). The loading plot clearly showed that plant height, panicle length, panicle width, seed yield per plot, 100 seed weight, seed volume, bulk density, true density and seed size contributed traits towards diversity. In this study, concluded that significant diversity existed among mini core accessions of sorghum for the traits studied. Efficient exploitation of this diversity

is helpful in identifying sorghum parental lines for hybrid breeding program. Inter-crossing between accessions from diverse clusters will provide segregating sorghum progeny for yield; thereby, leading to the development of high-yielding varieties.

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### References

- Anonymous. World Agricultural production, 2019a, 21.
- Anonymous. World Agricultural production, 2019b, 21
- Abraha, T., Githiri, S. M. Kasili, R. Araia, W. and Nyende. A. B. (2015). "Genetic variation among sorghum (*Sorghum bicolor* (L.) Moench) landraces from eritrea under postflowering drought stress conditions." American Journal of Plant Sciences. 6 (09): 1410. doi:10.4236/ajps.2015.69141
- Ali, M. A., Jabran, K., Awan, S. I., Abbas, A., Zulkiffal, M., Acet, T. and Rehman, A. (2011). "Morpho-physiological diversity and its implications for improving drought tolerance in grain sorghum at different growth stages." Australian Journal of Crop Science. 5 (3): 311.
- Ayana, A. and Bekele, E. (1999). "Multivariate analysis of morphological variation in sorghum (*Sorghum bicolor* (L.) Moench) germplasm from ethiopia and eritrea." genetic resources and crop evolution. 46 (3): 273–84. doi:10.1023/A:1008657120946.
- Bello, D. (2004). "Genetic variability and inter-relationship of traits in local sorghum (*Sorghum bicolor* (L.) Moench) in adamawa state." unpublished M.Sc. Thesis, Yola: Federal University of Technology.
- Chikuta, S., Odong, T., Kabi, F. and Rubaihayo, P. (2015). "Phenotypic diversity of selected dual purpose forage and grain sorghum genotypes." American Journal of Experimental Agriculture 9: 6. doi:10.9734/AJEA/2015/20577.
- Cochran, W.C. and Cox, G.M. (1957). Experimental designs. John Wiley and Sons, N.Y. London, pp. 82-90.
- Das, L. V. (2000). Problems facing plant breeding. New Delhi: CBS.
- Jain, S.K. and Patel, P.R. (2013). Multivariate analyses in sorghum [*Sorghum bicolor* (L.) Moench] for fodder yield and their attributes. Agric. Sci. Digest. 33(3):215-218.
- Johnson, D. E. (2012). Applied multivariate methods for data analysis. New York: Duxbury Press.
- Kotal, B. D., Das, A. and Choudry, B. K. (2010). Genetic variability and association of characters in wheat (*Triticum aestivum*L.). Asian J. Crop Sci. 2:155–160.
- Mahalanobis, P.C. (1936). On the generalized distance in statistics. Proceedings of National Institute of Sciences, India. 2:49-55
- Mary, S. S. and Gopalan, A. (2006). "Dissection of genetic attributes yield traits of fodder cowpea in F<sub>3</sub> and F<sub>4</sub>." Journal Applications Sciences Researcher.2 (6): 805–08.
- Mohanraj, K., Gopalan, A. and Shanmuganathan, M. (2006). Genetic diversity in sorghum [*Sorghum bicolor* (L.) Moench].The Journal of Agricultural Sciences, 2 (2):06-11.
- Murty, B.R. and Arunachalam, V.(1966). The nature and divergence in relation to breeding systems in some crop plants. Indian Journal of Genetics. 22:66-80
- Rao, C.R. (1952). Advanced statistical methods in biometrical research. New

- York, USA. John Wiley and Sons Inc.
- Sanchez, A., Subudhi, P. Rosenow, D. and H. Nguyen, 2002. Mapping QTLs associated with drought resistance in sorghum (*Sorghum SW* (L.) Moench). *Plant molecular biology*, 48(5-6): 713-726
- Sonawane, M.N., and Patil. F.B. (1991). Genetic divergence in cowpea. *Journal of Maharashtra Agricultural University*. 6: 167-169.
- Swami, S.S., Chaudhary, S.B. and Kute, N.S. (2015). Genetic diversity studies in *rabi* sorghum [*Sorghum bicolor* (L.) Moench]. *Journal of Agricultural Research Technology*. 40(2): 203-207.
- Watson, J.W. and Eyzaguirre, P.P.B. (2002). Homegardens and in situ conservation of plant genetic resources in farming systems. Proceedings of the second international home gardens workshop, 17-19th July 2001, Witzenhausen, Federal Republic of Germany. International Plant Genetic Resources Institute, Rome, Italy.
- Yan, W. and Kang. M. S. (2003). *GGE Biplot Analysis: A graphical tool for breeders, Geneticists, and Agronomists*. Boca Raton, FL: CRC Press.

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